# ROLE OF ESTRADIOL IN THE DEVELOPMENT OF FIBROSIS IN CYSTIC SCLERODERMA (POSSIBLE SEX-LINKED PREDISPOSITION TO THE DISEASE)

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Most research workers now consider that the development of fibrosis, which gives systemic scleroderma (SSD) its specific features, is linked with increased biosynthesis of collagen proteins by fibroblasts [10, 13]. The fibroblasts of patients with SSD are characterized by an excessively high level of procollagen-mRNA of types I and III and by a coordinated increase in the synthesis of the corresponding proteins [6, 8]. The principal factors specifically affecting the synthesis of particular proteins are steroid hormones (SH).

It has been shown that synthesis of type I procollagen by fibroblasts is controlled by glucocorticoids (GC) and that the degree of depression of protein synthesis is determined by the concentration of GC receptors in the cells [12]. Estradiol ( $E_2$ ), one of the principal female sex SH, also has been shown to inhibit collagen synthesis in the skin [14]. According to data obtained on cultures of aortic cells  $E_2$  depresses the synthesis of type III procollagen [2].  $E_2$  receptors have been found in the skin of various animals [4, 5]. These observations indicate that collagen synthesis can be regarded as an  $E_2$ -dependent process, a conclusion which has attracted particular attention in connection with the stronger predisposition to SSD of women.

This paper gives the results of a study of the  $E_2$ -receptor apparatus and the character of action of the hormone on type III procollagen synthesis by cultures of skin fibroblasts from normal individuals and patients with SSD.

# EXPERIMENTAL METHOD

Cultures of fibroblasts (3rd-4th passages, stationary phase of growth), obtained from skin biopsy specimens from the forearm of 17 clinically healthy persons (one man, the rest women, ages from 20 to 36 years) and 17 patients with SSD (all women, from 16 to 46 years old, duration of the disease 1-6 years). The disease followed an acute course in two patients, subacute in seven, and chronic in eight.

The cell cultures used in all the experiments were obtained from patients undergoing treatment in the Institute of Rheumatology, Academy of Medical Sciences of the USSR. Culture of the fibroblasts and determination of specific binding of  $E_2$  by whole cells (in suspension) were carried out by the method in [1]. Specific binding of  $E_2$  by a homogenate of fibroblasts was determined [5] with a final  $E_2$  concentration in the incubation medium of 80 nM.

In experiments using Scatchard plot analysis fibroblasts were cultured until a monolayer was formed (7th-8th days of culture) in flasks with a bottom surface area of about 5 cm<sup>2</sup>. The culture medium was changed 24 h before the experiment for similar medium, but containing 1% serum. Each determination was made on three or four flasks. The cells were incubated, using a culture medium (with 1% serum) containing 10-100 nM 2,4,6,7-(n)<sup>3</sup>H-E<sub>2</sub>, with specific radioactivity of 140 Ci/mmole (Amersham, England). In parallel tests a 200-fold excess of unlabeled hormone was added Incubation proceeded for 30 min at 37°C with constant shaking. The process was arrested by rapid removal of the incubation medium. The cell layer was washed 3 times

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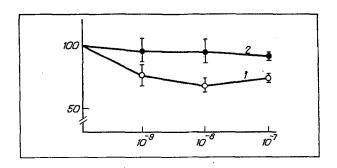


Fig. 1. Effect of  $E_2$  on production of type III procollagen by skin fibroblasts of normal individuals (n = 4) and patients with SSD (n = 5). Abscissa, concentration of  $E_2$  (in M); ordinate, production of type III procollagen (in % of control).

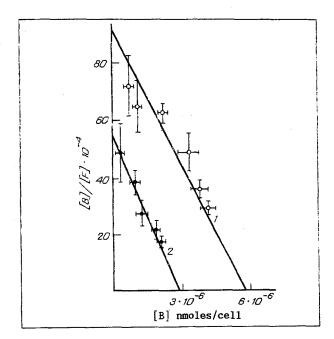


Fig. 2. Scatchard plot for binding of  ${}^{3}\text{H-E}_{2}$  by skin fibroblasts from normal individuals (1) and patients with SSD (2).

to remove radioactive unbound hormone with physiological saline The cells were next lysed by the addition of 200  $\mu$ l of 0.4% Na deoxycholate and the radioactivity of the samples was determined in the following scintillation mixture: methylcellosolve B 4 ml, toluene, containing 0.5% 2,5-diphenyloxazole, 5 ml.

The results of determination of specific binding of  $E_2$  in the homogenate, whole cells, or monolayer culture were comparable: under normal conditions (irrespective of the method used) specific binding of  $E_2$  amounted to 15-30% of the total.

The quantity of type III procollagen synthesized by fibroblasts was assessed on the basis of accumulation of the aminoterminal propertide type III procollagen, whose concentration was measured by radioimmunoassay [11].  $E_2$  was added to the cultures 24 h before the experiment in concentrations of  $10^{-9}$ ,  $10^{-8}$ , or  $10^{-7}$  M. The equivalent quantity of ethanol was added to the control cultures. Assistance with the technique and with some of the experiments was given by the Finnish research workers L. and J. Risteli, based in the Department of Clinical and Medical Chemistry, University of Oulu (Finland).

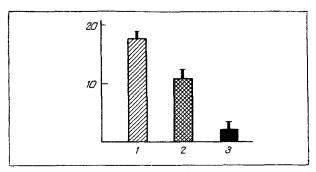


Fig. 3. Specific binding of  $E_2$  by skin fibroblasts from patients with SSD depending on type of course of the disease. 1) Normal individuals (n = 16), 2) chronic SSD (n = 5), 3) acute and subacute SSD (n = 8) Ordinate, binding of  $E_2$  (in % of total).

## **EXPERIMENTAL RESULTS**

Data on the effect of  $E_2$  on production of type III procollagen by fibroblasts from normal individuals and patients with SSD are given in Fig. 1. Under normal conditions  $E_2$  depressed production of type III procollagen: if the cells were cultured in the presence of  $10^{-9}$  M  $E_2$  the concentration of type III procollagen in the culture medium was  $77.5 \pm 10.3\%$  of the control (n = 4), in the presence of  $10^{-8}$  M  $E_2$  it was  $68.6 \pm 6.6\%$  (n = 4), and in the presence of  $10^{-7}$  M  $E_2$  it was  $80.0 \pm 3.1\%$  (n = 3). Under these circumstances, inhibition of type III procollagen production was observed in all the strains studied, and in the presence of all three concentrations of the hormone. Incidentally, these results are in agreement with data in the literature indicating that  $E_2$  inhibits synthesis of type III procollagen by vascular cells in culture [2].

In SSD the effect of the hormone was virtually undetectable: during cell culture in the presence of  $10^{-9}$  M  $E_2$  the concentration of type III procollagen in the culture medium was  $96.2 \pm 9.9\%$  (n = 4) of the control (i.e., the same as during culture of the cells without  $E_2$ ), in the presence of  $10^{-8}$   $E_2$  it was  $97.3 \pm 10.4\%$  (n = 5), and in the presence of  $10^{-7}$  M  $E_2$  it was 92.5% (n = 2). Inhibition of type III procollagen production under normal conditions compared with that in SSD was significant (p < 0.05) if concentrations of  $10^{-8}$  and  $10^{-7}$  M  $E_2$  were used.

The results are thus evidence that under normal conditions synthesis of type III procollagen by fibroblasts is an  $E_2$ -dependent process. Weakening of the effect of the hormone on this parameter can be regarded as an important factor in the hyperproduction of type III collagen and the development of fibrosis in SSD.

It is now considered that the physiological responses of the cells to the action of SH are mediated by receptors. In particular, it has been shown that the quantity of type I procollagen synthesized by fibroblasts depends on the concentration of GC receptors [12] and that activation of the synthesis of this protein in SSD is evidently due to reduction of receptor binding of GC in this disease [9].

Experiments accordingly were carried out to determine the number of  $E_2$  receptors in skin fibroblasts of healthy donors and patients with SSD. Scatchard plot analysis (Fig. 2) shows that the number of  $E_2$  receptors in the patients' fibroblasts was 1.96 times (p < 0.02) less than in normal subjects. The affinity of the hormone for the receptor was unchanged in SSD, however, as shown by the value of the binding constant  $K_b$ , which was  $8 \cdot 10^{10}$  M<sup>-1</sup> in normal individuals and  $8.5 \cdot 10^{10}$  M<sup>-1</sup> in SSD Consequently, in the modern view, weakening of the effect of  $E_2$  on the basic function of fibroblasts in patients with SSD can evidently be attributed to a decrease in the number of receptors of the hormone in these cells.

The results of four experiments on normal material and three on material from SSD patients are given in Fig. 2. Determination of binding of the hormone, using only one point on the Scatchard plot (specific binding) made it possible to compare receptor binding of  $E_2$  in a large group of healthy individuals (n = 16) and of patients with SSD (n = 13). In eight strains from SSD patients receptor binding was absent or was at a very low level, and in the remaining five cases it did not exceed 50-60% of the normal level, A low level of specific binding of  $E_2$  (13% of normal, p < 0.01) was observed in strains from patients with a less favorable type of course of the disease (acute or subacute). Binding of the hormone in cells from patients with chronic SSD (Fig. 3) was found in all strains tested, and was on average depressed by about half (p < 0.02). These results show that the degree of decrease in the number of  $E_2$  receptors corresponds to the severity of the pathological process. The

possibility cannot be ruled out that an unfavorable course of the disease process may be due to loss of hormone receptors not only by skin fibroblasts, but also by cells of other types of connective tissue.

Considering that smooth-muscle cells and vascular endothelial cells synthesize relatively large quantities of type III procollagen [3], control of the synthesis of this isoform by  $E_2$  is evidently particularly important. Damage to blood vessel walls, characteristic of SSD, may be connected with weakening of the inhibitory effect of  $E_2$  (coupled with loss of receptors), resulting in excessive accumulation of type III collagen in blood vessel walls. In addition, an excess of type III collagen may be a factor in the development of vascular lesions, for ability to induce platelet aggregation is a feature of this isoform to a greater degree than of others [7].

This investigation demonstrated the importance of the principal female sex hormone  $E_2$  in the regulation of fibroblast function.  $E_2$  is known to be present also in the male, but in a much lower concentration. Dependence of tissues in the female on  $E_2$  must evidently be greater than in the male. Lowering of the blood  $E_2$  level in women with SSD was found in our previous investigation [1], and weakening of the regulatory action of  $E_2$  on connective tissue cells ought to be manifested mainly in women, and this to a certain extent explains the linkage between sex and the incidence of SSD.

In conclusion, the authors are grateful to L and J. Risteli for help with the technique of determination of type III procollagen, and for providing reagents, and also to Professor K. Kivirikko for interest in the work and providing facilities for it in the Department of Clinical and Medical Chemistry, University of Oulu (Finland).

## LITERATURE CITED

- 1. M. D. Grozdova, A. F Panasyuk, L. N. Kashnikova, et al., Ter. Arkh., No. 5, 39 (1984).
- 2. J. Beldekas, B. Smith, et al., Biochemistry, 22, 2162 (1981).
- 3. J. Burke, G. Balian, R. Ross, and P. Bernstein, Biochemistry, 16, 3243 (1977).
- 4. U. Eppenberger and S. L. Hsia, J. Biol. Chem., 247, 5463 (1972).
- 5. H. Fleming, R. Blumenthal, and E. Gurpide, J. Steroid Biochem., 20, No. 1, 5 (1984).
- 6. P. N. Graves, I. K. Weiss, J. C. Perlish, et al., J. Invest. Derm., 80, 130 (1983).
- 7. J. Hugues, F. Herion, B. Husgens, and C. N. Lapiere, Thromb. Res., 9, 223 (1976).
- 8. S. A. Jimenez, G. Feldman, R. J. Baskey, and R. Bienkowski, Biochem. J., 237, 837 (1986).
- 9. L. N. Kashnikova, A. F. Panasyuk, and G. V. Yaroshchuk, Fourth Prague Rheumatological Symposium, Prague (1989), p. 62.
- 10. C. Le Roy, J. Clin. Invest., 54, 880 (1974).
- 11. O. Niemalä, L. Risteli, E. A. Sotaniemi, and J. Risteli, Clin. Chim. Acta, 124, 39 (1982).
- 12. A. Oikarinen, H. Oikarinen, E. M. L. Tan, and J. Uitto, Acta Derm.-Venereol. (Stockholm), 67, 106 (1987).
- 13. G. P. Rodnan, Immunological Diseases, ed. by M. Samter, Boston (1978), p. 1109.
- 14. T. Takeda, Y. Susuki, and C. S. Yao, Acta Path. Jpn., 25, 135 (1975).
- 15. M. Uzuka, K. Nakajima, and J. Mori, Biokhim. Biophys. Acta, 544, 329 (1978).